

Structure and transcription of the *Helicoverpa armigera* densovirus (HaDV2) genome and its expression strategy in LD652 cells

by Xu, P., Graham, R.I., Wilson, K. and Wu, K.

Copyright, Publisher and Additional Information: Publishers version distributed under the terms of the Creative Commons Attribution License:

<http://creativecommons.org/licenses/by/4.0/>

DOI: 10.1007/s10658-016-1113-5



RESEARCH

Open Access



Structure and transcription of the *Helicoverpa armigera* densovirus (HaDV2) genome and its expression strategy in LD652 cells

Pengjun Xu^{1,2}, Robert I. Graham³, Kenneth Wilson⁴ and Kongming Wu^{1*}

Abstract

Background: Densoviruses (DVs) are highly pathogenic to their hosts. However, we previously reported a mutualistic DV (HaDV2). Very little was known about the characteristics of this virus, so herein we undertook a series of experiments to explore the molecular biology of HaDV2 further.

Results: Phylogenetic analysis showed that HaDV2 was similar to members of the genus *Iteradensovirus*. However, compared to current members of the genus *Iteradensovirus*, the sequence identity of HaDV2 is less than 44% at the nucleotide-level, and lower than 36, 28 and 19% at the amino-acid-level of VP, NS1 and NS2 proteins, respectively. Moreover, NS1 and NS2 proteins from HaDV2 were smaller than those from other iteradensoviruses due to their shorter N-terminal sequences. Two transcripts of about 2.2 kb coding for the NS proteins and the VP proteins were identified by Northern Blot and RACE analysis. Using specific anti-NS1 and anti-NS2 antibodies, Western Blot analysis revealed a 78 kDa and a 48 kDa protein, respectively. Finally, the localization of both NS1 and NS2 proteins within the cell nucleus was determined by using Green Fluorescent Protein (GFP) labelling.

Conclusion: The genome organization, terminal hairpin structure, transcription and expression strategies as well as the mutualistic relationship with its host, suggested that HaDV2 was a novel member of the genus *Iteradensovirus* within the subfamily *Densovirinae*.

Keywords: Densovirus, HaDV2, Expression, Transcription

Background

The subfamily of *Densovirinae* within the family *Parvoviridae* is a group of small (18–26 nm diameter), non-enveloped, icosahedral viruses containing a linear single-stranded DNA genome ranging between 4 and 6 kb with characteristic terminal hairpins [1–3]. Members of this subfamily typically produce “cellular dense nucleosis” pathogenesis in their hosts, hence, they are commonly termed densoviruses (DVs) [4–8]. Since the first identification of a densovirus in the greater wax moth *Galleria mellonella* [9], DVs have been isolated from many arthropods, including species from six insect orders

(Lepidoptera, Diptera, Orthoptera, Dictyoptera, Odonata and Hemiptera) and decapod crustaceans (shrimps and crabs) [10–12].

To date, many DVs have been identified and sequenced. Unlike vertebrate parvoviruses, which all exhibit a monosense organization of their genome with nonstructural protein (NS) and structural protein (VP) open reading frames (ORFs) located on the same strand, arthropod DVs possess two types of genomes: monosense and ambisense [13–18]. Previously, the taxonomy of DVs was ambiguous, which was based on the organization of coding sequences, as well as genome size, terminal hairpin structure, gene expression strategy and host range [19]. Under the proposal of the International Committee on Taxonomy of Viruses (ICTV), Cotmore et al. [2] reconstructed the taxonomy of the family *Parvoviridae* in which DVs were

* Correspondence: kmwu@ippcaas.cn

¹State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, No. 2 West Yuan Ming Yuan Road, Beijing 100193, People's Republic of China
Full list of author information is available at the end of the article

classified into five distinct genera: *Ambidensovirus*, *Brevi-densovirus*, *Iteradensovirus*, *Hepandensovirus* and *Penstyl-densovirus* according to phylogenetic analysis and sequence homology.

DVs are highly pathogenic viruses to their hosts, and have been documented as being transmitted both horizontally and vertically [7, 9, 16]. Traditionally, these properties have captured the interest of many researchers investigating the potential application of DVs as biopesticides for biological control of insect pests or vectors for transgenic insects [20–26]. However, we previously reported a novel DV displaying a mutualistic interaction with its host (*Helicoverpa armigera*), and named this virus HaDV2 (previously named HaDNV-1) to distinguish it from the HaDV1 reported by El-Far et al. [27–29]. In this current study, we report the genome organization, transcription and expression strategies of the virus HaDV2.

Methods

Insect cell culture and transfection

Lymantria dispar LD652 cells, a gift from Central China Normal University in Wuhan (China) [30], were cultured in Grace's insect medium containing 10% fetal bovine serum (FBS) and 1% Penicillin-Streptomycin (Invitrogen, Grand Island, NY, USA) at 28 °C. Purified plasmid (100 ng) containing the ORFs of NS1 and NS2 with TIANpure Mini Plasmid Kit (TIANGEN, Beijing, China) was transfected into cells using Cellfectin® II Reagent as recommended (Invitrogen). The luciferase activities were determined using Luciferase Assay System (Promega, Madison, WI, USA).

Sequence analysis

Identity and alignment of the nucleotide and amino acid sequences was calculated using CLUSTAL W software [31]. The ORFs were identified using the ORF Finder (<http://www.ncbi.nlm.nih.gov/orffinder/>). Neighbor-joining trees with Poisson-corrected distances for the DV nucleotide sequences and the amino acid sequences of (NS1, NS2 and VP ORFs) were constructed using CLUSTAL W software and MEGA6.0 software [32].

Amplification of the stem-loop structure of HaDV2 genome by inverse PCR

The viral DNA was extracted from purified virus particles using TIANamp Genomic DNA Kit (TIANGEN). According to the reported genome sequence of HaDV2 (GenBank accession No.: HQ613271), three forward primers near the 3' end (DVF1 [nt 4576–4595], DVF2 [nt 3891–3910], DVF3 [nt 4343–4362]), and two reverse primers near the 5' end (DVR1 [nt 1038–1057] and DVR2 [nt 832–889]) were designed according to the genome sequence of HaDV2 (Additional file 1: Table S1). PCR reactions were performed using TransTaq DNA

Polymerase High Fidelity (TransGen, Beijing, China) and extracted viral DNA as a template. The PCR program was as follows: 30 s at 94 °C, 30 s at 57 °C, and 60 s at 72 °C for 40 cycles.

Mapping of the transcripts by 5'/3' RACE and Northern blot

The 5' and 3' ends of the HaDV2 transcripts were amplified using the SMART RACE cDNA Amplification Kit (Clontech, CA, USA), according to the manufacturer's instructions. cDNA was synthesized by RT-PCR from the total RNA of migrating cotton bollworms infected by HaDV2 using primers NS3F1/UPM for the 3' end, NS5R1/UPM and NS5R2/UPM for the 5' end of the NS genes, 3 F1/UPM for the 3' end, VP5R1/UPM and VP5R2/UPM for the 5' end of the VP genes, respectively (Additional file 1: Table S1). RNA (30 µg total) from insects infected by HaDV2 were separated on 1.1% formaldehyde agarose gels using MOPS buffer and blotted onto a positively charged nylon membrane (Roche, USA). Northern blot hybridization was performed using DIG-labeled probes (DIG DNA Labeling and Detection Kit, Roche, USA), according to the manufacturer's instruction. A 543 bp NS probe (1073–1615 nt) and a 420 bp VP probe (4081–4500 nt) were amplified by PCR with primers pairs NSF/NSR and VPF/VPR (Additional file 1: Table S1) using 30 cycles on a thermocycler as follows: 30 s at 95 °C, 30 s at 50 °C, and 30 s at 72 °C.

Antibody production

Using the predicted amino acid sequences gained from the earlier experiments in this study, two polypeptides were synthesized to raise polyclonal antisera in rabbits: CWDRAEFLRKYRKKVN and CDIGKSELWAPSVNPT for NS1 and NS2 proteins, respectively. The polypeptide of NS1, NS2 or VP was each emulsified with an equal volume of Freund's complete adjuvant for the first injection and incomplete adjuvant for subsequent injections. Antisera were obtained by injecting an adult rabbit subcutaneously with 500 µg polypeptide, followed by three additional injections of 300 µg polypeptide at 20 days intervals. The serum was purified and stored at –70 °C. The titer of the antisera was measured using ELISA as described by Liu et al. [33].

NS protein expression and subcellular localization

To characterize the expression of HaDV2 NS proteins in insect cells, we constructed two plasmids. Firstly, the HaDV2 NS promoter was amplified by primers NSPF/NSPR (Additional file 1: Table S1), digested with restriction endonuclease *KpnI/HindII* and cloned into a luciferase reporter vector pGL-3 Basic (Promega). This created the pNSP-Luc plasmid, in which the luciferase gene was under the control of the HaDV2 promoter.

Secondly, the complete ORFs of HaDV2 were amplified and cloned into the pEASY-T Cloning Vector (TransGen) to create pHaDENV-T with the primers HDVF1/HDVR1, HDVF2/HDVR2, HDVF3/HDVR3 (Additional file 1: Table S1), and using restriction endonucleases *Sac*II, *Afl*III and *Bsp*1407I. The resulting plasmid pHaDENV-T contained the whole genome of HaDV2 except for the hairpin structure. LD652 cells transfected with pHaDENV-T plasmid were then analyzed using 12% SDS-PAGE and transferred onto PVDF membranes. The membranes were blocked with dry skimmed milk (5%) and incubated in PBST buffer (PBS containing 0.1% Tween-20) containing polyclonal antibodies (anti-VP, anti-NS1 or anti-NS2) (1:5000, 1 h) and then a horseradish peroxidase (HRP)-conjugated secondary antibody (ZSGB-BIO, China) (1:20000, 1 h). The blots were revealed using the Easysee Western Blot Kit (Transgen).

To investigate the subcellular localization of these proteins, the NS1 and NS2 ORFs were amplified with primers NS1LF/NS1LR and NS2LF/NS2LR (Additional file 1: Table S1), then cloned into plasmid pIE-Atg6-GFP [34] by exchanging BmAtg6 with these two ORFs to create NS1-GFP and NS2-GFP constructs, respectively. The constructs were then transfected into LD652 cells and the NS-GFP fusion proteins were examined by fluorescent microscopy 24 h post-transfection. The IE2-GFP plasmid which was constructed by inserting the OpIE2 promoter of the pIZ-V5/His (Invitrogen) into the plasmid pEGFP-N1 digested by restriction enzymes *Bgl* II and *Sac* I was used as control and obtained as a gift from Dr. Liu's lab in Central China Normal University (China).

Results

Nucleotide sequences based analysis of HaDV2 genome organization

The size of virus particles of HaDV2 is about 20 nm in diameter and it possesses a monosegment genome about 5 kb. The sequencing analysis of the HaDV2 genome reported in our previous study showed that it differed considerably from other known DVs (Additional file 1: Figure S1) [28]. HaDV2 contained three large ORFs on the same strand ORF1 (1260 nt in length) encoded the putative NS2 polypeptide of 419 amino acids with a predicted molecular mass of 48 kDa and a theoretical isoelectric point (pI) of 7.10. ORF2 (2010 nt in length) encoded the putative NS1 protein of 669 amino acids with a predicted molecular mass of 78 kDa and a pI of 5.87. Alignment of the HaDV2 NS1 amino acid sequence with three other iteradenosoviruses indicated that HaDV2 NS1 protein shared two functional domains. Firstly, amino acids between aa 258–311 contained the two highly conserved replication initiator motifs involved in initiation and termination of rolling circle replication; and secondly, amino acids from aa 514–635 contained

the NTP-binding and helicase domains typical of the NS1 polypeptide. Alignment analysis indicated that the amino acid sequences of NS1 and NS2 from the HaDV2 were shorter than those from closely related members of the genus *Iteradenosovirus* (Additional file 1: Figure S2). ORF3 (1980 nt in length) encoded a putative VP protein of 659 amino acids with a molecular mass of 75 kDa and a pI of 7.13. The highly conserved phospholipase A2 domain located at amino acid positions aa 2–56 was also found in the VP ORF of HaDV2.

Tree-based analysis

Neighbor-joining trees were constructed using the whole genome sequence of HaDV2 as well as the putative amino acid sequences of NS1, NS2 and VP ORFs. Thirty-one DVs from other arthropods that had either the complete genome or the full coding sequence were included in the analysis (Fig. 1). The trees revealed five main branches: branch 1 included all known DV members with an ambisense genome in the genus *Ambidenosovirus*; branch 2 included members of the genus *Iteradenosovirus* and HaDV2; branch 3 included members of the genus *Hepandenosovirus*; branch 4 included members of the genus *Penstylidenosovirus*; and branch 5 consisted of members of the genus *Brevidenosovirus* (Fig. 1a). The branches of the tree constructed with amino acid sequences of VP was similar to that of the nucleotide sequence tree (Fig. 1b). The trees constructed with amino acid sequences of NS1 and NS2 differed considerably from the trees described above (see Fig. 1c, d). Although the four trees were not identical, they all indicated that the HaDV2 was most closely related to the members of the genus *Iteradenosovirus*. We also reconstructed the trees using Maximum likelihood method and the trees showed similar topology with the NJ trees except for NS2-tree (Additional file 1: Figure S3). Alignment of the nucleotide sequence and amino acid sequences (of VP, NS1 and NS2) indicated that sequence identities between viruses within the genus *Iteradenosovirus* exceed 58, 71, 35 and 28%, respectively; and that the identities between HaDV2 and members of the genus *Iteradenosovirus* are no more than 44, 36, 28 and 19%, respectively.

Determination of the stem-loop structure of HaDV2 genome by inverse PCR

Sequence analysis indicated that the HaDV2 terminal ends could form a stem-loop structure by a reverse complementary sequence (ITRs) near the two ends (Fig. 2a). To further confirm this prediction, PCR successfully amplified segments with forward primers near the 3' end and reverse primers near the 5' end. Sequence alignment indicated that all the amplified sequences were consistent with those of the HaDV2 except for the stem region located at ITRs (Fig. 2b).

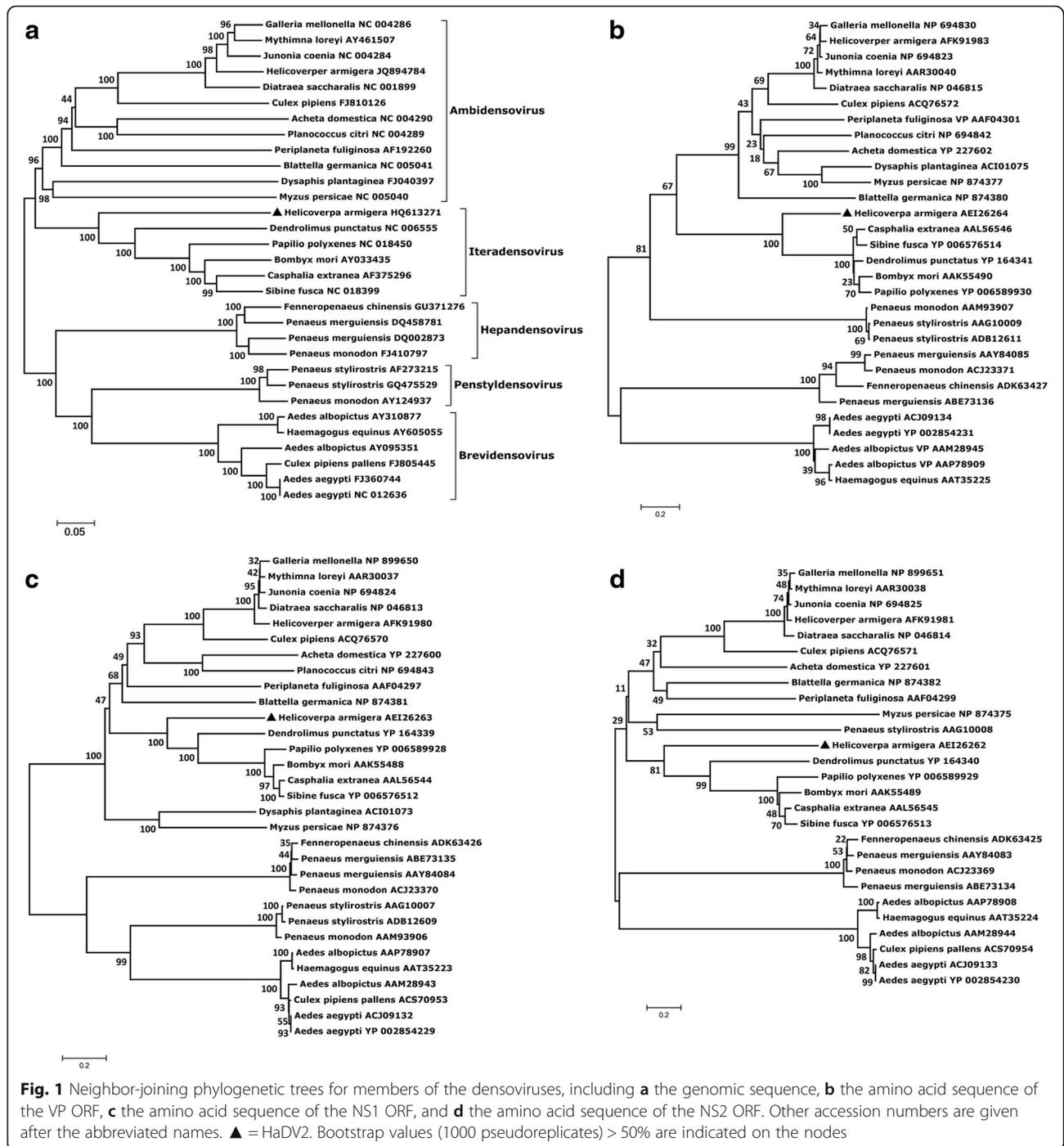


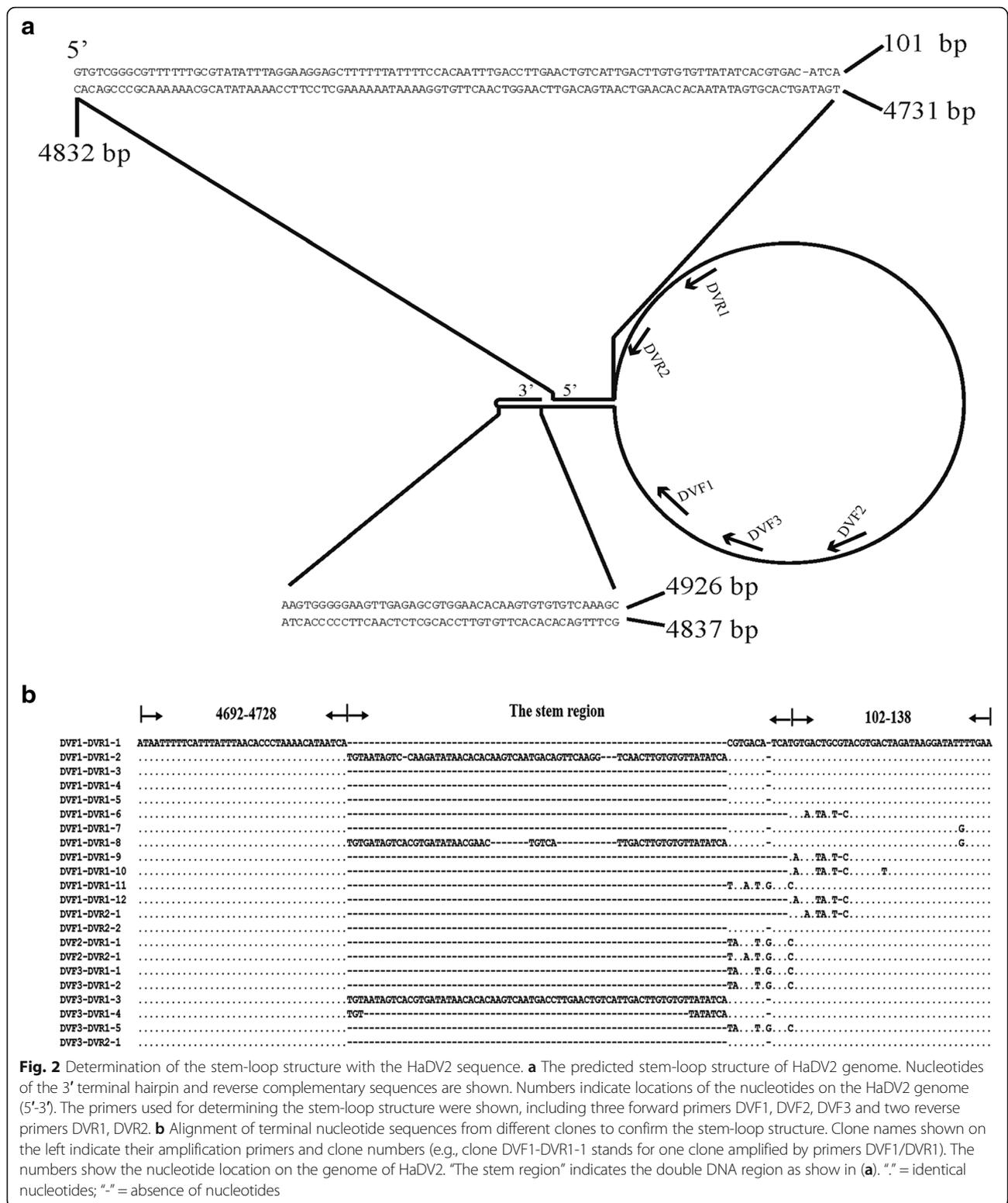
Fig. 1 Neighbor-joining phylogenetic trees for members of the densovirus, including **a** the genomic sequence, **b** the amino acid sequence of the VP ORF, **c** the amino acid sequence of the NS1 ORF, and **d** the amino acid sequence of the NS2 ORF. Other accession numbers are given after the abbreviated names. ▲ = HaDV2. Bootstrap values (1000 pseudoreplicates) > 50% are indicated on the nodes

Transcript analysis of HaDV2

Using 5' and 3' RACE primers, two transcription initiation sites (TISs) (at positions nt 307 (Fig. 3b) and nt 2516 (Fig. 3c)) and two transcription termination sites (TTSs) (at positions nt 2516 (Fig. 3d) and nt 4688 (Fig. 3e)) were determined. The TTS for the NS genes occurred at nt 2516, with canonical poly(A) addition sites (AATAAA) located 16 nucleotides upstream from

the TTS (Fig. 4d). The VP TTS was located at nt 4688, with canonical poly(A) addition sites situated 18 nucleotides upstream from the TTS (Fig. 4e). The transcript of 2209 bp ending at nt 2516 may encode the NS1 and NS2 proteins. The transcripts of 2173 bp ending at nt 4688 may encode the VP proteins (Fig. 4a).

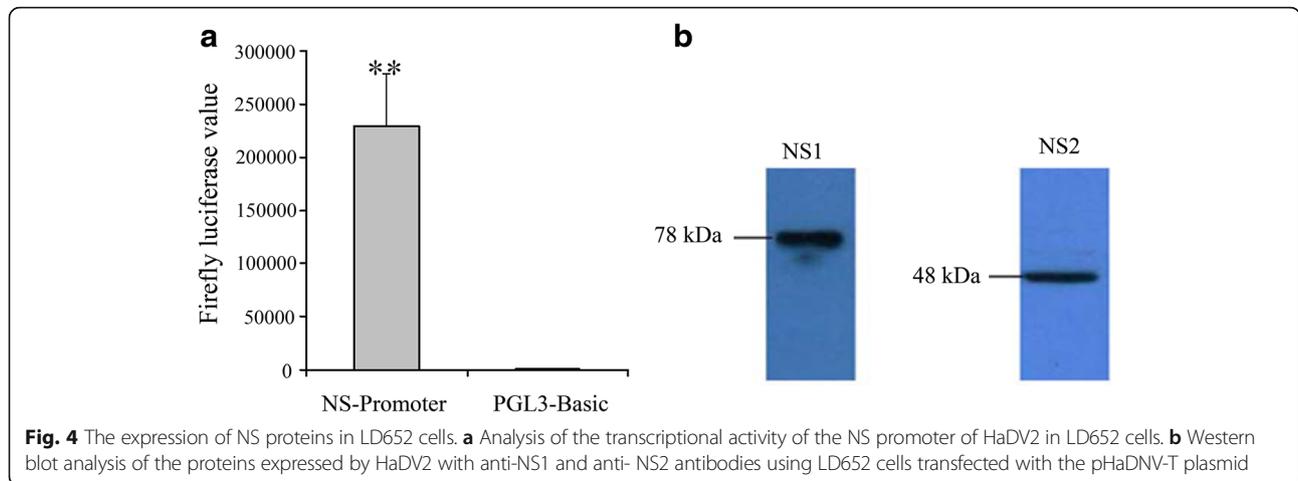
Northern Blot analysis was undertaken to determine the size and relative abundance of the transcripts for each of



the three viral ORFs. Hybridization of total RNA isolated from infected insects revealed one band of 2.2 kb when using the NS probe; and one band of 2.2 kb when using the VP probe (Additional file 1: Figure S4a, b).

The expression and subcellular localization of NS1 and NS2 proteins of HaDV2 in LD652 cells

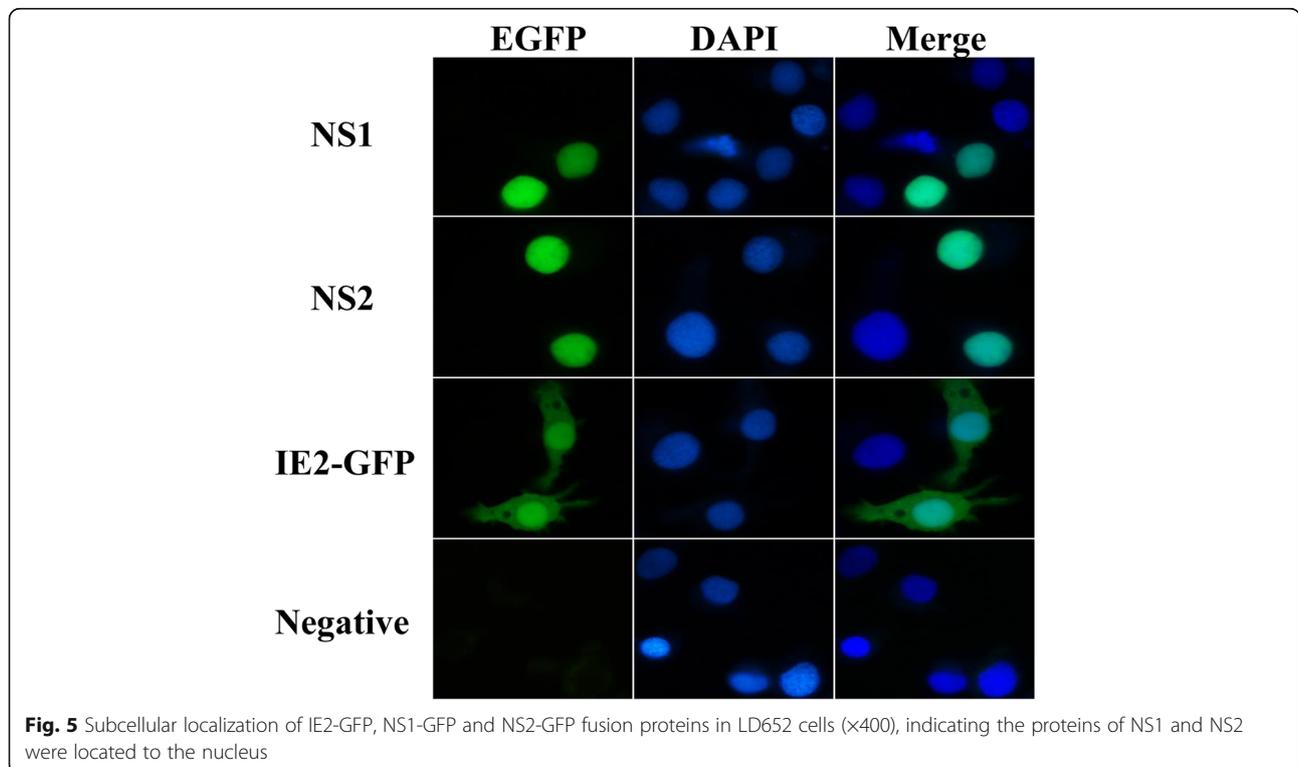
The expression of HaDV2 NS proteins was undertaken in *Lymantria dispar* LD652 cells. To determine the



In this current study, we determined the molecular biology of the HaDV2 virus, namely through examining its genome structure and ORF transcription and expression strategy. Based on our results, HaDV2 was a novel member of genus *Iteradenovirus*, with new features differing from other members from this genus, such as an ITR of 101 nt at both termini, a single 90 nts hairpin structure at the 3' end and the first ORF encoding NS2 protein [17, 28, 35–40].

Phylogenetic analysis using both nucleotide and amino acid sequences showed that HaDV2 was clustered within

the genus *Iteradenovirus*. The sequence identities of the viral DNA and the amino acid identities for VP, NS1 and NS2 ORFs among members of the genus *Iteradenovirus* exceed 58, 71, 35 and 28%, respectively. However, the sequence identities between HaDV2 and the current members of the genus *Iteradenovirus* are no more than 44, 36, 28 and 19%, respectively. Thus, although the HaDV2 was clustered with *Iteradenovirus*, it differs considerably from the other iteradenoviruses and appears to have a different function as described previously [29].



Although phylogenetic analysis indicated that HaDV2 was clustered with members of the genus *Iteradensovirus*, the NS1 and NS2 proteins of the HaDV2 are smaller than those of other *Iteradensovirus* (more than 753 and 451 amino acids, respectively) [17, 28, 35–40]. Are they functionally expressed as predicted? We used Western Blot analysis of transfected LD652 cells using anti-NS1 and anti-NS2 to show that the NS1 and NS2 proteins were 78 kDa and 48 kDa, respectively, consistent with the predicted size of the protein. NS proteins are a pivotal factor for viral transcription and replication as well as pathogenicity. The replication of DVs occurred in the nucleus of their hosts [19]. Therefore, the NS proteins of DVs should be located in the nucleus by nuclear localization signal (NLS) as reported by Yu et al. [40]. To further investigate whether the NS proteins of the HaDV2 localized within the nucleus of their hosts (as those of other DVs), NS1 and NS2 proteins were expressed in LD652 cells using the recombinant plasmid NS1-GFP and NS2-GFP. The result indicated that the NS1 and NS2 proteins were completely located in the nucleus, suggesting that they possess a common function and could possibly play a role in the novel interactions between HaDV2 and its host. The experiments with the NS proteins were carried out by transient expression in LD652 cells, which were not the virus's original host. It is acknowledged that this expression may not reflect the real role of HaDV2-NS promoter and how it works in the natural host.

Transcriptional patterns are diverse among the DVs. For example, JcDV, *Galleria mellonella densovirus* (GmDV) and *Mythimna loreyi densovirus* (MIDV) all have one transcript for the VP gene and two transcripts for the NS genes (the larger one for NS1 and the smaller one for NS2), in which the ORFs of NS1 and NS2 share a common TTS [15, 30, 41]. Meanwhile, the transcripts of CpDV, *Periplaneta fuliginosa densovirus* (PfdV) and *Myzus persicae nicotianae densovirus* (MpnDV) arise from alternative splicing [13, 42, 43]. The first ORFs of all known iteradensoviruses encode NS1 protein and the ORFs of NS2 are completely included in the ORFs of NS1 [40]. However, the first ORF of HaDV2 encodes NS2, which may impact gene expression of NS2 compared to NS1. In addition, the NS1 and NS2 of other iteradensoviruses were translated from different transcripts and the TIS of NS1 was found to start 2–26 nt upstream of the start codon [40]. Unexpectedly, our results suggested that the NS1 and NS2 of HaDV2 translated from the same transcript which started 63 nt upstream of the start codon of NS2. Surprisingly, although we provide evidence of the activity of the NS promoter, we failed to find the TATA-box upstream of the TIS of NS. Two TATA-box like sequences were located at nts 313 and 335 upstream of the start codon of NS1 and

NS2; suggesting HaDV2, maybe like brevidensoviruses, has overlapping NS gene promoters responsible for different transcript starts and dictating the relative transcription rates of these transcripts. However, one of the two transcripts was in great excess, making it difficult to detect both transcripts by RACE. Like other DVs, the VP transcripts of HaDV2 had short-untranslated regions, located at 5 nts upstream of the start codon of VP.

Conclusion

We report a novel densovirus, assigned as HaDV2, which differs from the other DVs in its genome organization, terminal hairpin structure, and transcription and expression strategies. Taken together with the unique mutualistic relationship previously described between HaDV2 and its host [29], this strongly indicates that HaDV2 is a novel member within the genus *Iteradensovirus*.

Additional file

Additional file 1: Table S1. Primers used in this study. **Figure S1.** The virus particles and its genome organization. (a) Electron micrograph of HaDENV-1 viruses purified from adult *Helicoverpa armigera* negatively stained with uranyl acetate (×200000). Bar, 100 nm. (b) Agarose gel electrophoresis (1%) of the extracted HaDENV-1 DNA. Lane 1 = DNA from the HaDENV-1, Lane 2 = Marker. (c) The putative ORFs of HaDENV-1. The plus strand contains three large ORFs: ORF1, ORF2 and ORF3, which encode NS2, NS1 and VP proteins, respectively. (d) Hairpin structure in the 3' terminus of HaDENV-1 predicted by the QuickFOLD program. The numbers in bracket stand for the start and stop nucleotides of the hairpin on the HaDENV-1 genome. **Figure S2.** Alignment of amino acid sequences of NS1 (a) and NS2 (b) of HaDENV-1 with the ones of members from *Iteradensovirus*. HaDENV = *Helicoverpa armigera* densovirus 1 (accession number: HQ613271), BmDENV = *Bombyx mori* densovirus 1 (AY033435), CeDENV = *Casphalia extranea* densovirus (AF375296), DpDENV = *Dendrolimus punctatus* densovirus (NC_006555). **Figure S3.** The maximum-likelihood tree for members of the densoviruses, including (a) the genomic sequence with GTR + G + I model, (b) the amino acid sequence of the VP ORF with LG + G model, (c) the amino acid sequence of the NS1 ORF with LG + G + I model, and (d) the amino acid sequence of the NS2 ORF with JTT + G model. "▲" represents the sequence of HaDV2. Bootstrap values (1000 pseudoreplicates) > 50% are indicated on the nodes. **Figure S4.** Northern blot analysis of the HaDV2 transcripts showed two bands of 2.2 kb with the NS and the VP probe, respectively. **Figure S5.** Dose-responses of anti-NS1, anti-NS2 and anti-VP antibodies using ELISA. (DOC 2203 kb)

Abbreviations

DV: Densovirus; ICTV: The international committee on taxonomy of viruses; NS: Nonstructural protein; ORF: Open reading frame; pI: Theoretical isoelectric point; VP: Structural protein

Acknowledgements

We would like to thank Dr. Jie Wang (Tobacco Research Institute, Chinese Academy of Agricultural Sciences, Qingdao, P.R. China) and Dr. Peng Xu (Central China Normal University, Wuhan, P.R. China) for providing suggestions and repairing figures.

Funding

This research was supported by Science Fund for Creative Research Groups of the National Science Foundation of China (Grant No. 31321004), the National Natural Science Foundation of China (Grant No. 31401752).

Authors' contributions

KWu and PX conceived the study. PX performed the experiments. KWu, PX, RIG and KWu wrote the manuscript. All of the authors critically reviewed and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval and consent to participate

No permit was required to collect the tested the virus in insect. Sampling did not involve regulated, endangered or protected species.

Author details

¹State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, No. 2 West Yuan Ming Yuan Road, Beijing 100193, People's Republic of China. ²Tobacco Research Institute, Chinese Academy of Agricultural Sciences, No. 11 Ke Yuan Jing Si Road, Qingdao 266101, People's Republic of China. ³Crop and Environment Sciences, Harper Adams University, Newport TF10 8NB, UK. ⁴Lancaster Environment Centre, Lancaster University, Lancaster LA1 4YQ, UK.

Received: 10 October 2016 Accepted: 18 January 2017

Published online: 07 February 2017

References

- Berns KI, Bergoin M, Bloom M, Lederman M, Muzyczka N, Siegl G, et al. Parvoviridae: Vth report of international committee on taxonomy of viruses. *Arch Virol.* 1995;10(Suppl):169–78.
- Cotmore SF, Agbandje-McKenna M, Chiorini JA, Mukha DV, Pintel DJ, Qiu J, et al. The family Parvoviridae. *Arch Virol.* 2014;159:1239–47.
- Dhar AK, Robles-Sikisaka R, Saksmerprome V, Lakshman DK. Biology, genome organization, and evolution of parvoviruses in marine shrimp. *Adv Virus Res.* 2014;89:85–139.
- Amargier A, Vago C, Meynardier G. Histopathological study of a new type of viral disease demonstrated in the Lepidoptera *Galleria mellonella*. *Arch Gesamte Virusforsch.* 1965;15:659–67.
- Hu Y, Zheng J, Lizuka T, Bando H. A densovirus newly isolated from the smoky-brown cockroach *Periplaneta fuliginosa*. *Arch Virol.* 1994;138:365–72.
- O'Neill SL, Kittayapong P, Braig HR, Andreadis TG, Gonzalez JP, Tesh RB. Insect densoviruses may be widespread in mosquito cell lines. *J Gen Virol.* 1995;76:2067–74.
- Rivers CF, Longworth JF. A nonoccluded virus of *Junonia coenia* (Nymphalidae: Lepidoptera). *J Invertebr Pathol.* 1972;20:369–70.
- Vago C, Quiot JM, Luciani J. Infection of lepidopteran tissue culture with the purified virus of the densovirus. *C R Acad Sci Hebd Seances Acad Sci D.* 1966;263:799–800.
- Meynardier G, Vago C, Atger P. Virose d'un type inhabituel chez le lepidoptere *Galleria mellonella* L. *Rev Zool Agric Appl.* 1964;63:207–9.
- Fediere G. Epidemiology and pathology of *Densovirinae*. *Contrib Microbiol.* 2000;4:1–11.
- Safeena MP, Tyagi A, Rai P, Karunasagar I. Complete nucleic acid sequence of *Penaeus monodon densovirus* (PmDENV) from India. *Virus Res.* 2010;150:1–11.
- van Munster M, Dullemans AM, Verbeek M, van den Heuvel JF, Reinbold C, Brault V, et al. Characterization of a new densovirus infecting the green peach aphid *Myzus persicae*. *J Invertebr Pathol.* 2003;84:6–14.
- Baquerizo-Audiot E, Abd-Alla A, Jousset FX, Cousserans F, Tijssen P, Bergoin M. Structure and expression strategy of the genome of *Culex pipiens densovirus*, a mosquito densovirus with an ambisense organization. *J Virol.* 2009;83:6863–73.
- Guo H, Zhang J, Hu Y. Complete sequence and organization of *Periplaneta fuliginosa densovirus* genome. *Acta Virol.* 2000;44:315–22.
- Tijssen P, Li Y, El-Far M, Szelei J, Letarte M, Zadori Z. Organization and expression strategy of the ambisense genome of densovirus of *Galleria mellonella*. *J Virol.* 2003;77:10357–65.
- van Munster M, Dullemans AM, Verbeek M, van den Heuvel JF, Reinbold C, Brault V, et al. A new virus infecting *Myzus persicae* has a genome organization similar to the species of the genus *Densovirus*. *J Gen Virol.* 2003;84:165–72.
- Wang J, Zhang J, Jiang H, Liu C, Yi F, Hu Y. Nucleotide sequence and genomic organization of a newly isolated densovirus infecting *Dendrolimus punctatus*. *J Gen Virol.* 2005;86:2169–73.
- Zhai YG, Lv XJ, Sun XH, Fu SH, Gong ZD, Fen Y, et al. Isolation and characterization of the full coding sequence of a novel densovirus from the mosquito *Culex pipiens pallens*. *J Gen Virol.* 2008;89:195–9.
- Bergoin M, Tijssen P. Molecular biology of *Densovirinae*. *Contrib Microbiol.* 2000;4:12–32.
- Bossin H, Fournier P, Royer C, Barry P, Cerutti P, Gimenez S, et al. *Junonia coenia* densovirus-based vectors for stable transgene expression in Sf9 cells: influence of the densovirus sequences on genomic integration. *J Virol.* 2003;77:11060–71.
- Corsini J, Afanasiev B, Maxwell IH, Carlson JO. Autonomous parvovirus and densovirus gene vectors. *Adv Virus Res.* 1996;47:303–51.
- Giraud C, Devauchelle G, Bergoin M. The densovirus of *Junonia coenia* (Jc DNV) as an insect cell expression vector. *Virology.* 1992;186:207–18.
- Hu L, Zhang L, Shen C, Lu J, Zhang J, Hu Y. The densovirus of *Periplaneta fuliginosa* (PFDNV) as an insect vector for persistent foreign gene expression in vivo. *Biochem Biophys Res Commun.* 2007;358:976–82.
- Jiang H, Zhang J, Wang J, Yang B, Liu C, Lu J, et al. Genetic engineering of *Periplaneta fuliginosa densovirus* as an improved biopesticide. *Arch Virol.* 2007;152:383–94.
- Mutuel D, Ravallec M, Chabi B, Multeau C, Salmon JM, Fournier P, et al. Pathogenesis of *Junonia coenia densovirus* in *Spodoptera frugiperda*: a route of infection that leads to hypoxia. *Virology.* 2010;403:137–44.
- Suzuki Y, Niu G, Hughes GL, Rasgon JL. A viral over-expression system for the major malaria mosquito *Anopheles gambiae*. *Sci Rep.* 2014;4:5127.
- El-Far M, Szelei J, Yu Q, Fediere G, Bergoin M, Tijssen P. Organization of the ambisense genome of the *Helicoverpa armigera densovirus*. *J Virol.* 2012;86:7024.
- Xu P, Cheng P, Liu Z, Li Y, Murphy RW, Wu K. Complete genome sequence of a monosense densovirus infecting the cotton bollworm, *Helicoverpa armigera*. *J Virol.* 2012;86:10909.
- Xu P, Liu Y, Graham RI, Wilson K, Wu K. Densovirus is a mutualistic symbiont of a global crop pest (*Helicoverpa armigera*) and protects against a baculovirus and Bt biopesticide. *PLoS Pathog.* 2014;10:e1004490.
- Wang Y, Abd-Alla AM, Bossin H, Li Y, Bergoin M. Analysis of the transcription strategy of the *Junonia coenia densovirus* (JcDENV) genome. *Virus Res.* 2013;174:101–7.
- Thompson JD, Higgins DG, Gibson TJ. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* 1994;22:4673–80.
- Tamura K, Stecher G, Peterson D, Filipiński A, Kumar S. MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol.* 2013;30:2725–9.
- Liu C, Gao Y, Ning C, Wu K, Oppert B, Guo Y. Antisera-mediated in vivo reduction of Cry1Ac toxicity in *Helicoverpa armigera*. *J Insect Physiol.* 2010;56:718–24.
- Wei W, Gai Z, Ai H, Wu W, Yang Y, Peng J, et al. Baculovirus infection triggers a shift from amino acid starvation-induced autophagy to apoptosis. *PLoS One.* 2012;7:e37457.
- Fediere G, Li Y, Zadori Z, Szelei J, Tijssen P. Genome organization of *Casphalia extranea densovirus*, a new iteravirus. *Virology.* 2002;292:299–308.
- Li Y, Zadori Z, Bando H, Dubuc R, Fediere G, Szelei J, et al. Genome organization of the densovirus from *Bombyx mori* (BmDENV-1) and enzyme activity of its capsid. *J Gen Virol.* 2001;82:2821–5.
- Yu Q, Fediere G, Abd-Alla A, Bergoin M, Tijssen P. Iteravirus-like genome organization of a densovirus from *Sibine fusca* Stoll. *J Virol.* 2012;86:8897–8.
- Yu Q, Hajek AE, Bergoin M, Tijssen P. *Papilio polyxenes densovirus* has an iteravirus-like genome organization. *J Virol.* 2012;86:9534–5.
- Yu Q, Tijssen P. *Iteradensovirus* from the monarch butterfly, *Danaus plexippus plexippus*. *Genome Announc.* 2014;2:e00321–14.
- Yu Q, Tijssen P. Gene expression of five different iteradensoviruses: *Bombyx mori densovirus*, *Casphalia extranea densovirus*, *Papilio polyxenes densovirus*, *Sibine fusca densovirus*, and *Danaus plexippus densovirus*. *J Virol.* 2014;88:12152–7.

41. Fediere G, El-Far M, Li Y, Bergoin M, Tijssen P. Expression strategy of densovirus from *Mythimna loreyi*. *Virology*. 2004;320:181–9.
42. Yamagishi J, Hu Y, Zheng J, Bando H. Genome organization and mRNA structure of *Periplaneta fuliginosa* densovirus imply alternative splicing involvement in viral gene expression. *Arch Virol*. 1999;144:2111–24.
43. Tang S, Song X, Xue L, Wang X, Wang X, Xu P, et al. Characterization and distribution analysis of a densovirus infecting *Myzus persicae nicotianae* (Hemiptera: Aphididae). *J Econ Entomol*. 2016;109:580–7.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

